Congenital Disorder of Glycosylation Ia: PMM2 Gene Sequencing

**Test Code:** SPMM2  
**Turnaround time:** 6 weeks  
**CPT Codes:** 81479 x1

### Condition Description

Congenital disorders of glycosylation (CDG) are a group of autosomal recessive genetic disorders caused by the alteration in synthesis and structure of protein and lipid glycosylation. In the past decade, over 30 genetic diseases have been identified that alter glycan synthesis, structure and ultimately the function of nearly all organ systems.

CDG type I (CDGI) disorders result from impaired synthesis of the incomplete lipid linked oligosaccharide (LLO) and/or its attachment to the growing polypeptide chain. CDG-Ia is the most common form reported, due to phosphomannomutase deficiency, an enzyme that converts mannose-6-phosphate to mannose-1-phosphate. CDG-Ib (phosphomannose isomerase, MPI deficiency) is the only known treatable form, by giving mannose orally. CDG type II (CDGII) includes defects in processing of N-glycans. Phenotypes of this disorder are extremely variable. Manifestations range from severe developmental delay and hypotonia with multiple organ system involvement beginning in infancy, to hypoglycemia and protein-losing enteropathy with normal development. Most subtypes have been described in only a few individuals, however, thus understanding of the phenotypes is limited.

The current diagnostic test for CDG is analysis of serum transferrin glycoforms, also called "transferrin isoforms analysis," or "carbohydrate-deficient transferrin analysis." If positive, this testing can be followed by DNA testing to identify mutations in the gene involved.

The typical clinical course of CDG Ia has been divided into an infantile multisystem stage, late-infantile and childhood ataxia-mental retardation stage, and adult stable disability stage. Recent reports have widened the phenotypic spectrum to include hydrops fetalis at the severe end and a mild neurologic phenotype in adults with multisystemic involvement at the mild end. Clinical presentation and course are highly variable, ranging from infants who die in the first year of life to mildly involved adults. Clinical presentations do, however, tend to be similar in siblings.

Because the presentation of this disorder is highly variable, the diagnosis should be considered in a child with development delay and hypotonia in combination with any of the following findings:

- Failure to thrive
- Hepatic dysfunction (elevated transaminases)
- Coagulopathy with low serum concentration of factors IX and XI, antithrombin, protein C, and/or protein S
- Hypothyroidism, hypogonadism
- Esotropia
- Pericardial effusion
- Abnormal subcutaneous fat pattern including increased suprapubic fat pad, skin dimpling, and inverted nipples or subcutaneous fat pads having a toughened, puffy, or uneven consistency
- Seizures
- Stroke-like episodes
- Osteopenia, scoliosis
- Cerebellar hypoplasia/atrophy and small brainstem

The diagnosis of CDG Ia should be considered in adolescents or adults with suggestive histories and any of the following findings:

- Cerebellar dysfunction (ataxia, dysarthria, dysmetria)
- Non-progressive cognitive impairment
- Stroke-like episodes
- Peripheral neuropathy with or without muscle wasting
- Absent puberty in females, small testes in males
- Retinitis pigmentosa
- Progressive scoliosis with truncal shortening
- Joint contractures

Confirmation of the diagnosis in a proband requires molecular genetic testing following the finding of a type I transferrin isoform pattern. Individuals with the clinical diagnosis of CDG Ia and biochemical diagnosis of PMM enzyme deficiency with normal transferrin glycosylation, however, have been reported. Mutations in the PMM2 gene (16p13.3-p13.2) cause CDG Ia, and in individuals with enzymatically proven CDG Ia, the mutation detection rate in PMM2 approaches 100%. The prevalence of CDG Ia could be as high as 1:20,000.

For patients with suspected CDG Ia, sequence analysis is recommended as the first step in mutation identification. For patients in whom mutations are not identified by full gene sequencing, deletion/duplication analysis is appropriate.


### References:

- GeneTests: Congenital Disorders of Glycosylation Type 1a

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## Genes

**PMM2**

### Indications

This test is indicated for:

- Individuals with a clinical/biochemical diagnosis consistent with CDG Ia.
- Carrier testing in individuals with a family history of CDG Ia.

### Methodology

PCR amplification of 8 exons contained in the PMM2 gene is performed on the patient's genomic DNA. Direct sequencing of amplification products is performed in both forward and reverse directions, using automated fluorescence dideoxy sequencing methods. The patient's gene sequences are then compared to a normal reference sequence. Sequence variations are classified as mutations, benign variants unrelated to disease, or variations of unknown clinical significance. Variants of unknown clinical significance may require further studies of the patient and/or family members. This assay does not interrogate the promoter region, deep intronic regions, or other regulatory elements, and does not detect large deletions.

### Detection

**Clinical Sensitivity:** In individuals with enzymatically proven CDG-1a, the mutation detection rate in PMM2 approaches 100%. Mutations in the promoter region, some mutations in the introns and other regulatory element mutations cannot be detected by this analysis. Large deletions will not be detected by this analysis. Results of molecular analysis should be interpreted in the context of the patient's biochemical phenotype.

**Analytical Sensitivity:** ~99%.

### Specimen Requirements

**Submit only 1 of the following specimen types**

**Type: Whole Blood (EDTA)**

**Specimen Requirements:**

EDTA (Purple Top)  
Infants and Young Children (2 years of age to 10 years old): 3-5 ml  
Older Children & Adults: 5-10 ml  
Autopsy: 2-3 ml unclotted cord or cardiac blood  

**Specimen Collection and Shipping:**  
Ship sample at room temperature for receipt at EGL within 24 hours of collection. Do not refrigerate or freeze.

**Type: DNA, Isolated**

**Specimen Requirements:**

Microtainer  
8µg  
Isolation using the Perkin Elmer™Chemagen™ Automated Extraction method or Qiagen™ Puregene kit for DNA extraction is recommended.

**Specimen Collection and Shipping:**  
Refrigerate until time of shipment in 100 ng/µL in TE buffer. Ship sample at room temperature with overnight delivery.

**Type: Saliva**

**Specimen Requirements:**

Oragene™ Saliva Collection Kit  
Orangene™ Saliva Collection Kit used according to manufacturer instructions. Please contact EGL for a Saliva Collection Kit for patients that cannot provide a blood sample.

**Specimen Collection and Shipping:**  
Please do not refrigerate or freeze saliva sample. Please store and ship at room temperature.

### Special Instructions

Please submit copies of diagnostic biochemical test results along with the sample. Contact the laboratory if further information is needed.

Sequence analysis is required before deletion/duplication analysis by targeted CGH array. If sequencing is performed outside of EGL Genetics, please submit a copy of the sequencing report with the test requisition.

### Related Tests

- Deletion/duplication analysis of the PMM2 gene by CGH array is available for those individuals in whom sequence analysis is negative.
- Analysis of other CDG genes is also available.
- Biochemical carbohydrate deficient transferrin analysis for CDGs is also available.
- Custom diagnostic mutation analysis (KM) is available to family members if mutations are identified by targeted mutation testing or sequencing analysis.
- Prenatal testing is available to adult couples who are confirmed carriers of mutations. Please contact the laboratory genetic counselor to discuss appropriate testing prior to collecting a prenatal specimen.